



THE UNIVERSITY *of* EDINBURGH

Edinburgh Research Explorer

Elucidation of the RamA Regulon in *Klebsiella pneumoniae* Reveals a Role in LPS Regulation

Citation for published version:

De Majumdar, S, Yu, J, Fookes, M, Mcateer, SP, Llobet, E, Finn, S, Spence, S, Monaghan, A, Kissenpfennig, A, Ingram, RJ, Bengoechea, J, Gally, DL, Fanning, S, Elborn, JS & Schneiders, T 2015, 'Elucidation of the RamA Regulon in *Klebsiella pneumoniae* Reveals a Role in LPS Regulation', *PLoS Pathogens*, vol. 11, no. 1, 1004627, pp. e1004627. <https://doi.org/10.1371/journal.ppat.1004627>

Digital Object Identifier (DOI):

[10.1371/journal.ppat.1004627](https://doi.org/10.1371/journal.ppat.1004627)

Link:

[Link to publication record in Edinburgh Research Explorer](#)

Document Version:

Publisher's PDF, also known as Version of record

Published In:

PLoS Pathogens

Publisher Rights Statement:

Copyright: © 2015 De Majumdar et al. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

General rights

Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy

The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.



RESEARCH ARTICLE

Elucidation of the RamA Regulon in *Klebsiella pneumoniae* Reveals a Role in LPS Regulation

Shyamasree De Majumdar^{1,2}, Jing Yu¹, Maria Fookes³, Sean P. McAteer⁴, Enrique Llobet⁵, Sarah Finn⁶, Shaun Spence¹, Avril Monaghan¹, Adrien Kissenpfennig¹, Rebecca J. Ingram¹, José Bengoechea^{1,5}, David L. Gally⁴, Séamus Fanning⁶, Joseph S. Elborn¹, Thamarai Schneiders^{1,2*}

1 Centre for Infection and Immunity, Belfast, United Kingdom, **2** Division of Pathway and Infection Medicine, Edinburgh, United Kingdom, **3** Wellcome Trust Sanger Institute, Wellcome Trust Genome Campus, Hinxton, Cambridge, United Kingdom, **4** Division of Immunity and Infection, The Roslin Institute and R(D) SVS, The University of Edinburgh, Easter Bush, Midlothian, United Kingdom, **5** Laboratory Microbial Pathogenesis, Fundació d'Investigació Sanitària de les Illes Balears (FISIB) Recinto Hospital Joan March, Bunyola, Spain, **6** UCD Centre for Molecular Innovation and Drug Discovery, School of Public Health, Physiotherapy & Population Science, University College Dublin, Dublin, Ireland

* thamarai.schneiders@ed.ac.uk



OPEN ACCESS

Citation: De Majumdar S, Yu J, Fookes M, McAteer SP, Llobet E, Finn S, et al. (2015) Elucidation of the RamA Regulon in *Klebsiella pneumoniae* Reveals a Role in LPS Regulation. PLoS Pathog 11(1): e1004627. doi:10.1371/journal.ppat.1004627

Editor: Alan Hauser, Northwestern University, UNITED STATES

Received: May 15, 2014

Accepted: December 14, 2014

Published: January 29, 2015

Copyright: © 2015 De Majumdar et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was supported by grants awarded to TS from the MRC New Investigator Grant G0601199, QUB start up funds, the ESCMID Research Grant Award and the Wellcome Trust grant 098051. DLG and SPM are supported by an Institute Strategic Programme Grant from the BBSRC. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

Abstract

Klebsiella pneumoniae is a significant human pathogen, in part due to high rates of multi-drug resistance. RamA is an intrinsic regulator in *K. pneumoniae* established to be important for the bacterial response to antimicrobial challenge; however, little is known about its possible wider regulatory role in this organism during infection. In this work, we demonstrate that RamA is a global transcriptional regulator that significantly perturbs the transcriptional landscape of *K. pneumoniae*, resulting in altered microbe-drug or microbe-host response. This is largely due to the direct regulation of 68 genes associated with a myriad of cellular functions. Importantly, RamA directly binds and activates the *lpxC*, *lpxL-2* and *lpxO* genes associated with lipid A biosynthesis, thus resulting in modifications within the lipid A moiety of the lipopolysaccharide. RamA-mediated alterations decrease susceptibility to colistin E, polymyxin B and human cationic antimicrobial peptide LL-37. Increased RamA levels reduce *K. pneumoniae* adhesion and uptake into macrophages, which is supported by *in vivo* infection studies, that demonstrate increased systemic dissemination of ramA overexpressing *K. pneumoniae*. These data establish that RamA-mediated regulation directly perturbs microbial surface properties, including lipid A biosynthesis, which facilitate evasion from the innate host response. This highlights RamA as a global regulator that confers pathoadaptive phenotypes with implications for our understanding of the pathogenesis of *Enterobacter*, *Salmonella* and *Citrobacter* spp. that express orthologous RamA proteins.

Author Summary

Bacteria can rapidly evolve under antibiotic pressure to develop resistance, which occurs when target genes mutate, or when resistance-encoding genes are transferred. Alternatively, microbes can simply alter the levels of intrinsic proteins that allow the organism to

“buy” time to resist antibiotic pressure. *Klebsiella pneumoniae* is a pathogen that causes significant blood stream or respiratory infections, but more importantly is a bacterium that is increasingly being reported as multidrug resistant. Our data demonstrate that RamA can trigger changes on the bacterial surface that allow *Klebsiella* to survive both antibiotic challenge, degradation by host immune peptides and resist phagocytosis. We demonstrate that the molecular basis of increased survival of *ramA* overexpressing *K. pneumoniae*, against host-derived factors is associated with RamA-driven alterations of the lipid A moiety of *Klebsiella* LPS. This modification is likely to be linked to *Klebsiella*’s ability to resist the host response so that it remains undetected by the immune system. The relevance of our work extends beyond RamA in *Klebsiella* as other pathogens such as *Enterobacter* spp and *Salmonella* spp. also produce this protein. Thus our overarching conclusion is that the intrinsic regulator, RamA perturbs host-microbe and microbe-drug interactions.

Introduction

The microbial response to antimicrobial challenge is multifactorial and can be conferred by a combination of extrinsic or intrinsic mechanisms. Those intrinsic mechanisms that confer pleiotropic phenotypes can provide a “stepping stone” to surmounting both the host or drug response. Intrinsic proteins such as the AraC-transcriptional proteins e.g. MarA [1], SoxS [2], Rob [3], RamA [4] and RarA [5], directly regulate genes linked to microbial permeability barriers which results in reduced susceptibility [6] to multiple antibiotic classes. The perturbation of the permeability barrier is identified as a critical step in the development and emergence of higher levels of resistance [7].

The regulatory proteins, typified by the MarA protein, are unique, as unlike other members of the AraC family, these proteins bind DNA as monomers [8], interact with RNA polymerase via a process of pre-recruitment [9] and generally confer reduced antimicrobial susceptibility [10]. Microarray analyses has highlighted the wider effects of increased MarA [1], SoxS [2], RamA [4, 11] and RarA [5] levels in modulating gene expression particularly of those genes linked to virulence. This is further supported by studies reporting that either the inhibition or deletion of these regulators [12] can impair the ability of *E. coli* to colonise and cause infection *in vivo* [13]. Taken together, it is evident that these AraC proteins can confer bifunctional phenotypes of reduced drug susceptibility and increased virulence, which facilitate pathogen survival. These findings firstly, underscore the relative importance of these factors in microbial survival and secondly, provide a rationale for the development of “Anti-virulence-type” inhibitors against these transcription proteins.

The *ramA* gene which encodes the RamA protein is found in *Klebsiella*, *Enterobacter* [14], *Salmonella* [15] and *Citrobacter* spp [16] where the genetic organisation of the *ram* locus is conserved in most organisms, with the exception of *Salmonella enterica* serovar Typhimurium (Fig. 1) which lacks *romA*, a putative metallo-beta-lactamase gene. The levels of both the *romA-ramA* genes are repressed at the transcriptional level by the TetR-type family regulator RamR, encoded by the *ramR* gene, which is divergently transcribed from the *romA-ramA* operon. In both *Klebsiella* and *Salmonella*, an increase in *ramA* expression can be mediated by inactivating mutations [16–18] or ligand mediated interactions [19] with the cognate repressor, RamR which binds to a highly conserved inverted repeat (atgagtgn₆cactcat) [20] overlapping the promoter region of the *romAramA* operon (Fig. 1). Mutations within the *ramR* gene in *K. pneumoniae* resulting in *ramA* overexpression were initially reported as a result of tigecycline

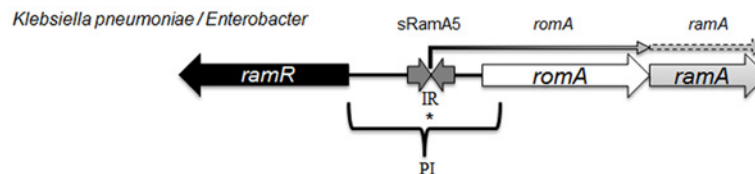


Figure 1. Organisation of the *ram* locus in *Klebsiella pneumoniae* / *Enterobacter* spp. In *K. pneumoniae* *romA* and *ramA* are usually co-transcribed from the TSS depicted with *, under the control of the PI promoter. RamR can bind the inverted repeat (atgagtgn₆cactcat), which in turn represses the transcription of both *romA* and *ramA*. Our analysis shows that the small regulatory RNA, sRamA5 and *romA* in *Klebsiella pneumoniae* share the TSS (depicted with *).

doi:10.1371/journal.ppat.1004627.g001

exposure [17, 21]. However, previous work evaluating clinical isolates that pre-date the use of tigecycline demonstrate that *ramA* overexpressing strains were already present within the nosocomial population of *K. pneumoniae*, suggesting a broader role for RamA mediated overexpression in antibiotic resistance [16]. Interestingly, studies evaluating the prevalence of *ramA*-mediated overexpression in clinical isolates of *K. pneumoniae* and *Salmonella* spp. indicate that these bacteria are more likely to overexpress *ramA* than *marA* or *soxS*, suggesting that elevated *ramA* levels may be more relevant to the development of antibiotic resistance in these organisms.

Several studies [4, 11] have addressed the scope of the RamA regulon in *Salmonella enterica* serovar Typhimurium using microarray profiling. These studies demonstrate that *ramA* overexpression results in reduced antimicrobial susceptibility due to the differential regulation of *acrAB* and *micF* genes, which consequently decrease OmpF levels. One study [4] suggests that genes linked to the Salmonella Pathogenicity Island (SPI-2) are also differentially expressed, leading to the initial observation that RamA may impact on *Salmonella*-specific virulence attributes. However this link was not corroborated in subsequent *in vivo* experiments. In *K. pneumoniae*, the wider impact of RamA-mediated regulation is not known. Despite the apparent similarities in genome structure, the microbial lifestyles of both *K. pneumoniae* [22] and *Salmonella* spp. differ. Importantly, the increasing multidrug resistance in *Klebsiella* spp. demands a thorough understanding of factors within this genus that contribute to the intrinsic microbial 'resistome' and survival under selective (host or drug) pressure. Therefore to define the broad effects of RamA-mediated expression on microbe-host and microbe-drug phenotypes we carried out transcriptome profiling using directional RNAseq with the wild type strain *K. pneumoniae* Ecl8 [23] and its isogenic derivatives Ecl8Δ*ramA* and Ecl8Δ*ramR*. Our key findings show the scope of RamA-mediated regulation significantly alters the transcriptional landscape of *K. pneumoniae*. This occurs by directly modulating the expression of different genes notably those associated with antimicrobial resistance and host-microbe interactions thereby resulting in the emergence of a less antibiotic susceptible and more virulent *K. pneumoniae*.

Results

Regulation of the *ram* Locus

The *ram* locus encodes a sRNA to maintain basal levels of *ramA* expression. RamR functions as the primary repressor of both *romA*-*ramA* expression in *K. pneumoniae* by binding the palindromic repeats of the IR element which flanks the TSS for *romA* at position -64T. *ramR*, itself, has two transcriptional start sites, located at the -83T and -167A positions where expression analyses using GFP fusions suggest that the primary promoter region for *ramR* transcription is located at the -83T start site (S1 and S2 Figs.). This site is also repressed 5-fold

more than the vector only control by *ramR* in trans indicating that like other TetR-type regulators, RamR expression is autoregulated (S2 Fig.).

Previous work in *Salmonella* has shown that the regulatory RNA, StyR3, can control expression at the *ram* locus [24]. Given the expansive role of *ramA* in gene regulation, we sought to determine whether the *K. pneumoniae* ortholog of StyR3, denoted as sRamA5, would function as co-regulator of *ramA* expression in *K. pneumoniae* to promote basal *ramA* levels. The lack of similarity within the intergenic regions located between the *ramR* and *romA-ramA* genes or *ramR* and *ramA* genes in *K. pneumoniae* and *Salmonella* spp. respectively, excluded the possibility of using sequence analyses to identify the StyR3 ortholog. Direct northern blot analyses of RNA derived from *K. pneumoniae* strain Ecl8 and its derivatives did not produce a detectable signal for the putative regulatory RNA, sRamA5. Thus in order to demonstrate the presence of sRamA5, we cloned the entire intergenic region flanked by the *ramR* and *romA* genes and the partial *romA* open reading frame into the TA cloning vector pGEMTeasy to generate pGEMsRamA5. Northern blot analyses derived from the expression of sRamA5 encoded on pGEMsRamA5, using gene specific probes for sRamA5 and *romA* ORF, demonstrate the presence of sRamA5 (~ 60nt) (shown in Fig. 2A). Notably, the sRamA5 specific probe also detected a further two RNA molecules (Fig. 2A, arrowed bands 1 and 2). These fragments, detected by both the sRamA5 and *romA* specific probe, possibly represent primary transcripts initiated from the common start site as determined by 5' RACE analyses for sRamA5 and *romA* (S1 Fig.). As expected the *romA* specific probe did not detect the 60nt sRamA5 molecule (Fig. 2A). Thus we surmise that sRamA5 and *romA* are co-transcribed into a primary RNA molecule, which undergoes further processing prior to excision proximal to the start of the *romA* gene, thereby producing sRamA5.

As a classical TetR-family protein, RamR-mediated repression of the *romA-ramA* locus is likely to be perturbed through ligand-mediated interactions; therefore we hypothesized that to function as a co-regulator of *romA-ramA* expression RamR would interact with sRamA5. RNA-EMSA (S1 Text) analyses demonstrate that RamR and sRamA5 form a complex, suggesting direct interaction of the RNA (sRamA5) with RamR (Fig. 2B). In order to ascertain whether the interaction of sRamA5 and RamR is attributable to the presence of the highly conserved IR sequence in the *ramR-romA* intergenic region (ATGAGTGcgctactCACTCAT) and thus, act as a competitor for RamR-pI binding, we performed EMSA analyses using the pI promoter, sRamA5 and RamR. Our results show a reduction in affinity of RamR to sRamA5 in the presence of excess pI promoter (Fig. 2B). In contrast, competition experiments with excess sRamA5 show no perturbation of the pI+RamR interaction, suggesting that RamR has a higher affinity for the pI promoter compared to sRamA5 (Fig. 2B).

Simultaneous qPCR measurements utilizing an LNA probe to assess sRamA5 levels demonstrate firstly, that the transcription levels of sRamA5 and *romA* are not linked as sRamA5 levels are decreased in contrast to elevated *romA* levels (Fig. 2C). This suggests that despite being transcribed from the same TSS, sRamA5 and *romA* are likely subject to different rates of degradation. Secondly, the stability of sRamA5 may be dependent on the presence of a functional RamR. In order to investigate the requirement for a functional RamR in sRamA5 stability, we determined both the *romA* and sRamA5 levels in Ecl8Δ*ramR* before and after complementation with *ramR* expressed in trans. As expected, our results show that the level of *romA* transcription was reduced (~ 30-fold) in Ecl8Δ*ramR*/pACramR compared to the plasmid only control (Ecl8Δ*ramR*/pACYC177) (Fig. 2D). In contrast, the levels of sRamA5 were found upregulated by ~ 2.8 fold in Ecl8Δ*ramR*/pACramR relative to the plasmid only control (Ecl8Δ*ramR*/pACYC177). Thus the increase in sRamA5 levels in the presence of a functional *ramR* supports our hypothesis that sRamA5 is stabilized by RamR. Our data also shows that sRamA5 does compete with pI for RamR binding, although this effect may be abrogated by the

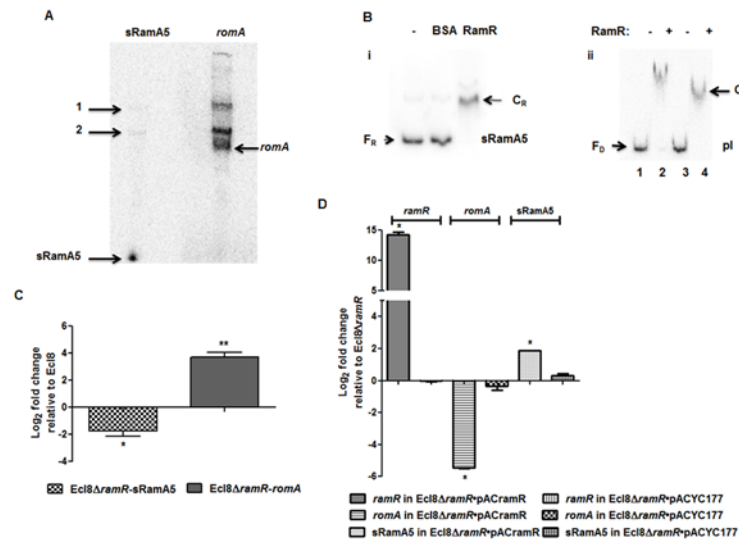


Figure 2. A: Northern blot analysis of sRamA5. 15 μg of total RNA extracted from Ecl8/pGEMTPI+romA were loaded into wells. The blots were either incubated with the ³²P-end labelled sRamA5 specific DNA probe or romA specific DNA probe. The bands pointed as 1 and 2 are primary transcripts for both the RNA codes for sRamA5 and romA. The band referred to as sRamA5 is specifically detected by the sRamA5 DNA probe, sized at around 60 nucleotides; the band referred to as romA was specifically detected by the romA DNA probe. **B: EMSA of RamR-sRamA5 or RamR-PI interaction** in the presence of sRamA5. (i). RamR-sRamA5 interaction. The concentrations of sRamA5 and RamR were 40 nM and 1 μM respectively. (ii). RamR-PI interaction in the absence/presence of sRamA5. Radioactive labeled pI was 2 nM from lane 1 to 4. RamR's concentrations from lane 1 to 4 were: 0, 2, 0, 2 μM. Cold sRamA5's concentrations from lane 1 to 4 were: 0, 0, 1, 1 μM. F_R = free RNA, C_R = RNA-protein complex, F_D = free DNA, C = RNA-DNA-protein complex. **C: qPCR for the level of romA and sRamA5's transcriptions** in Ecl8ΔramR. qPCR using LNA probe for determining the levels of sRamA5 transcription in Ecl8 and Ecl8ΔramR. Despite sharing the same TSS, the transcript levels of sRamA5 are not linked to romA levels, thereby reducing the likelihood of sRamA5 being a 5' untranslated region of romA. The log₂ fold changes in Ecl8ΔramR displayed in the bar chart are relative to their transcript levels in Ecl8. One-way ANOVA analyses ($P < 0.001$) were performed to demonstrate statistical significance. **D: qPCR assay for the level of ramR, romA and sRamA5** in Ecl8ΔramR pACYCramR and Ecl8ΔramR pACYC177. The log₂ fold changes in the two strains displayed in the bar chart are relative to their transcript levels in Ecl8ΔramR. sRamA5 levels are elevated in the presence of ramR, implying that RamR could stabilise the sRamA5 transcript. One-way ANOVA analyses ($P < 0.001$) were performed to demonstrate statistical significance.

doi:10.1371/journal.ppat.1004627.g002

higher relative affinity of RamR to the pI promoter (Fig. 2B(ii)). Therefore, we surmise that the physiological relevance of RamR-sRamA5 interaction supports the basal level of ramA transcription detected in the wild type *K. pneumoniae* Ecl8.

Describing the Transcriptional Landscape of *K. pneumoniae* Ecl8 and Its Isogenic Derivatives Ecl8ΔramA, Ecl8ΔramR

To determine the effect of altered RamA levels on the whole transcriptome of *K. pneumoniae* strain Ecl8, we quantitatively compared the transcriptomes of the three strains (Ecl8, Ecl8ΔramA, Ecl8ΔramR) using the Kolmogorov-Smirnov (K-S) 2-sample test (S3 Fig.) as described in the supplementary data [25]. As expected, the distribution curve of Ecl8 and Ecl8ΔramA were more similar to each other compared to that observed for Ecl8ΔramR, suggesting that under normal growth conditions the deletion of ramA is less likely to perturb the transcriptional landscape as opposed to when it is overexpressed. This supports the notion that ramA functions as a pleiotropic regulator of gene expression in *K. pneumoniae*.

In all three strains, the 16S and 23S rRNA genes showed the highest number of mapped reads consistent with the lack of depletion for ribosomal RNA. However, pairwise comparisons of the normalized basemean values associated with these ribosomal regions were not differentially expressed between Ecl8 and Ecl8 Δ ramR or Ecl8 and Ecl8 Δ ramA. The lack of differential ribosomal gene expression is contrary to previous observations in *Salmonella enterica* serovar Typhimurium [4]. Other non-ribosomal genes (e.g. *fusA_1* (encoding translation elongation factor G), *atpA* (producing ATP synthase F1, α subunit) and *aceE* (encoding a pyruvate dehydrogenase)) were also found to have significantly high basemean values relative to most other genes within the genome. The increased expression of these genes is perhaps not surprising as *atpA* is associated with aerobic growth and *aceE* catalyses the production of precursors to the TCA cycle.

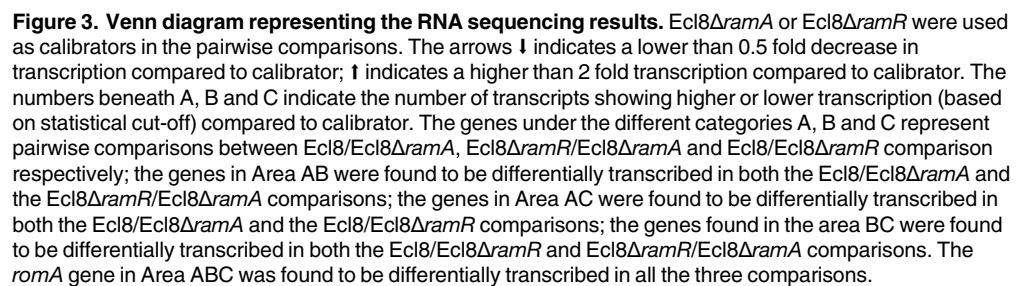
Potential regions of antisense transcription were also detected. However, in most cases, these regions appeared as antisense because of *in silico* errors in annotation or due to transcriptional noise from flanking genes within the chromosome. We did, however, identify antisense transcription, such as with BN373_16241 (producing an oxidoreductase) and BN373_02611, which were differentially expressed due to either elevated RamA levels or loss of the *ramA* gene (S4 Fig.). Coverage plots analyses indicate that the transcription associated with BN373_02611 may be associated with 3'UTR runoff transcription from the divergently transcribed *treBC* operon, in contrast to BN373_16241, which is upregulated when *ramA* was overexpressed and may be a "true" antisense RNA (S4 Fig.).

Genome analyses of *K. pneumoniae* strain Ecl8 [23] identified 11 unique predicted prophage genes encoding phage structural components (BN373_03311, BN373_09871, BN373_10091, BN373_14801, BN373_14811, BN373_14821, BN373_14841, BN373_14921, BN373_21511, BN373_37361, BN373_37371) which were not found to be differentially transcribed in the pairwise comparisons tested (Ecl8 vs Ecl8 Δ ramA, Ecl8 vs Ecl8 Δ ramR (S1 Table)). However, pairwise comparisons of Ecl8 Δ ramA and Ecl8 Δ ramR detected the differential expression of Ecl8-genome specific genes, BN373_33401, BN373_33411, which were repressed (~ 2 – 3 fold) in the *ramA* overexpressing strain Ecl8 Δ ramR (S2 Table). Of note, no differential gene expression was noted in the 233 plasmid-coding genes in the *ramA* null mutant or in the *ramA* over-expressor (Ecl8 Δ ramR) with respect to the wild type (Ecl8).

Defining the RamA Regulon

Transcriptome analyses underscores that perturbations in RamA levels can result in the differential expression of open reading frames, antisense transcripts and Ecl8-specific genes. As the majority of reads were mapped to open reading frames, the main focus of our analyses relates to the differential regulation of genes within *K. pneumoniae*. The RamA regulon in *K. pneumoniae* was identified by pairwise comparisons of Ecl8 Δ ramR versus Ecl8 (C) or Ecl8 Δ ramA (B). The pairwise comparisons of Ecl8 versus Ecl8 Δ ramA (A) (Fig. 3) indicate the cohort of genes (13) responsive to basal levels of RamA expression; the contrast between Ecl8 versus Ecl8 Δ ramR (35) specifies genes that are either affected by RamR or RamA, whereas the comparison between Ecl8 Δ ramR versus Ecl8 Δ ramA (77) identifies genes that largely react to altered RamA levels. As fewer genes are affected due to perturbations in *ramR* expression as opposed to RamA levels, we surmise that the majority of genes differentially expressed in our pairwise comparison (B) are associated with RamA-mediated regulation. Based on this assessment, the probable RamA regulon, Fig. 3, constitutes a total of 103 genes (as in genes in categories A, B, AB, BC, CA, ABC) (S2 Table). Of these, 68 genes were found to be activated and 35 were repressed (S2 Table) when levels of RamA is relatively higher.

Genes associated with RamA-mediated regulation were initially mapped to the COG (clusters of orthologous groups) database to explore their biological function. COG functional



classifications of the significantly differentially expressed genes reveal that RamA controls a myriad of cellular and metabolic processes (COG data presented in [S2 Table](#)). Generally, altered levels of RamA significantly modulate the expression of genes belonging to the COG functional group C (energy production and conversion). Specifically, when *ramA* is deleted, genes within the COG (G) (carbohydrate metabolism and transport) were also found to be differentially regulated. Pairwise comparison between *Ecl8ΔramR* versus *Ecl8* indicates that COG families associated with transcription (K) and inorganic ion transport and metabolism (P) were also affected. Additionally, when *ramA* levels are elevated genes associated with cell wall membrane and envelope biogenesis (M), transcription (K) and Function Unknown (FUN) categories were most differentially affected. Thus the resulting COG analyses also supports the observation where altered levels of RamA triggers a shift in gene functionality consistent with significant modulations in transcription patterns as predicted by the K-S test ([S3 Fig](#)).

PLOS Pathogens | DOI:10.1371/journal.ppat.1004627 January 29, 2015

such as *acrAB*, *oqxAB* and *yrbB-F*, which is consistent with phenotypes linked to multidrug resistance [27] and susceptibility to toxic small molecules, which is associated with alterations in the lipid symmetry of the cell wall [28]. However, the pairwise comparisons for *Ecl8* and *Ecl8-ΔramA* also suggest that basal levels of RamA are sufficient to trigger the upregulation of genes such as the trehalose transporter operon *treBC* and the ribose ABC transporter, *rbsACB*. Uniquely, genes associated with biofilm formation (*hha-ybaJ* encodes a toxin-antitoxin system) and lipid A biosynthesis BN373_10601 (encodes lipid A biosynthesis lauroyl acyltransferase, *lpxL_2*) and the related dioxygenase protein encoding gene *lpxO* (BN373_36331) were also found to be upregulated by RamA.

A total of 51 genes were found to be downregulated. As expected, *ompF* was significantly repressed in the *ramA* overexpresser (*Ecl8ΔramR*) (Fig. 3) in addition to genes encoding the nitrate reductases (*narGHJI* operon and *nirD*), BN373_05601 encoding the LysR-type transcriptional regulator, elongation factor EF2 and the riboflavin synthase encoding gene *ribH* were also found to be significantly downregulated in the *ramA* overexpresser (*Ecl8ΔramR*).

Only a subset of those differentially regulated genes was chosen for validation using qPCR. As expected, both the *romA* and *ramA* genes were found to show 5.25- \log_2 fold and 14.5- \log_2 fold increase in *Ecl8ΔramR* respectively compared to *Ecl8ΔramA* (S5A Fig.). When the activated genes (with the exception of *romA*, *ramA*) were assessed, increased expression of the following genes was noted (Fig. 4A): *tolC* (4.8- \log_2 fold), *acrA* (4.6- \log_2 fold), *yhbW* (1.8- \log_2 fold),

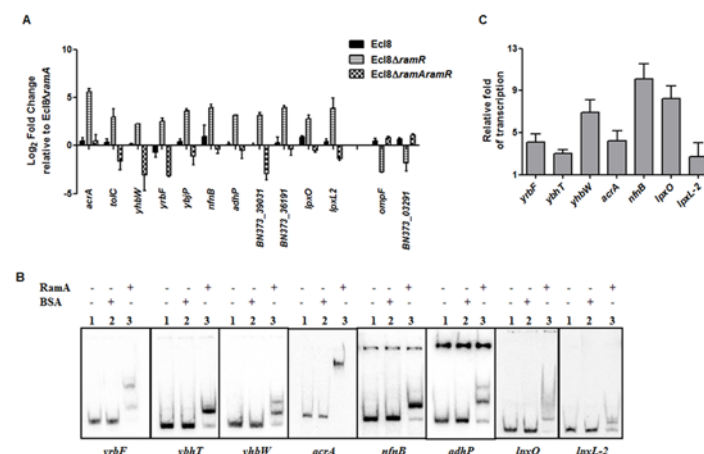


Figure 4. A: Quantitative real-time RT-PCR validation of differentially expressed genes in *Ecl8ΔramR*.

All qPCR experiments were performed as outlined in [materials and methods](#). Expression levels were normalized to 16S levels, and fold change values were generated by calibrating against *Ecl8ΔramA*. Genes designated BN373_36191, BN373_39031, BN373_03291 encode a putative membrane protein, oxidoreductase family and conserved hypothetical protein respectively. All data is a mean of 3 experiments. **B:** Electrophoretic Mobility Shift Assay (EMSA) using purified RamA protein. Following PCR amplification, each promoter region was end-labelled with ^{32}P - γ ATP. Purified RamA (200 nM) and the different labelled DNA probes (2 nM) were incubated on ice. All reactions were performed on ice prior to electrophoresis on 7.5% native gel. Lane 1 of each panel indicates the labelled DNA probe only, Lane 2 is the BSA control and Lane 3 contains RamA+DNA. **C:** Transcription *in vitro* assay of different promoters using the purified RamA protein. The test DNA (2 nM- *yrbF*, *yhbT*, *yhbW*, *acrA*, *nfnB*, *lpxO* and *lpxC*) with the control template (*gnd*) were incubated for transcription *in vitro* [^{32}P]α-UTP with (+) or without (-) 200 nM purified RamA. Samples were fractionated by polyacrylamide/urea gel electrophoresis prior to drying and exposure to the phosphorimager. Relative fold increase was determined using densitometric analysis as described previously [55], by first normalizing all test transcription levels to the control promoter (*gnd*) prior to comparison to the no protein control. Statistics was done using One way ANOVA (P value < 0.05) where transcription levels were found to be statistically significant in the presence of purified RamA compared to the no protein control.

doi:10.1371/journal.ppat.1004627.g004

yrbC (2.8- log₂ fold), *nfnB* (3.3- log₂ fold), *ybjP* (3.95- log₂ fold), *adhP* (3.2- log₂ fold), BN373_36191 (encodes putative membrane protein, 2.95- log₂ fold), BN373_39031 (encodes oxidoreductase, aldo/keto reductase family, 1.5- log₂ fold), *lpxO* (2.8- log₂ fold) and *lpxL-2* (3.6- log₂ fold). As expected, the levels of the *ompF* (4.2- log₂ fold down) and BN373_03291 (encodes conserved hypothetical protein, 1.1- log₂ fold down) were also downregulated (Fig. 4A).

Direct Regulation by RamA

In order to determine if some of these differentially expressed genes were under the direct or indirect control of RamA, we performed both EMSA and *in vitro* transcription (IVT) using purified recombinant RamA protein. The EMSA results show that RamA directly binds the *yrbF*, *ybhT*, *yhbW*, *acrA*, *nfnB*, *adhP*, *lpxO* and *lpxL-2* promoters (Fig. 4B). Of note, our controls, showed no shift in the presence of the test promoters (Fig. 4B).

We then determined whether RamA would directly regulate the different promoters identified. By performing IVT experiments, we initially tested the effects of the RamA protein against the *acrAB* promoter to ascertain if RamA would function correctly as a transcriptional activator. As expected, the purified recombinant RamA activated the *acrAB* promoter directly (Fig. 4C) thereby confirming the biological activity of the purified RamA protein. Subsequently, we assessed the test promoters identified by the EMSA in our IVT assays. The results show that RamA upregulates *yrbF* (4-fold), *ybhT* (3-fold), *yhbW* (6.9-fold), *acrA* (4-fold), *nfnB* (10-fold), *lpxO* (8-fold) and *lpxL-2* (~3-fold) (Fig. 4C). Thus purified recombinant RamA alone can directly activate the expression of these promoters *in vitro*.

Functional Relevance of RamA-Mediated Overexpression

RamA regulates genes involved in lipid A biosynthesis. Having established that purified RamA directly binds and activates the expression of *lpxL-2* and *lpxO* gene promoters (Fig. 4B and 4C), we sought to determine whether RamA could regulate other genes associated with the lipid A biosynthetic pathway. The lipid A biosynthetic pathway is governed by nine enzymes encoded by *lpxA*, *lpxC*, *lpxD*, *lpxB*, *lpxK*, *lpxI*, *lpxM* and *lpxO* genes [29]. Gene expression analyses using qPCR showed that with the exception of *lpxC*, none of the other *lpx* genes showed significant differential expression in Ecl8Δ*ramR* in comparison to Ecl8 or Ecl8Δ*ramA* (Fig. 5A). We then chose to assess whether RamA would directly interact with the *lpxC* and *lpxK* promoter regions. Subsequent EMSA analyses demonstrate that RamA directly interacts with the *lpxC* but not the *lpxK* promoter (Fig. 5Bi) and increased *lpxC* transcription (9-fold) in the presence of purified RamA and RNA polymerase (Fig. 5Bii). Previous work has shown that the control of lipid A biosynthetic genes is mediated by the PhoPQ or PmrAB systems [30]. Further interrogation of the transcriptome data and subsequent qPCR analyses shows that the levels for *phoP* and *pmrA* levels remained unchanged in *K. pneumoniae* Ecl8, Ecl8Δ*ramA* and Ecl8Δ*ramR*. Thus the differential modulation of the *lpxO*, *lpxC* and *lpxL-2* genes is directly linked to increased RamA levels.

To ascertain whether RamA-mediated transcriptional activation of *lpxC*, *lpxL-2* and *lpxO* would actually result in modifications within the lipid A moiety, we performed MALDI TOF mass spectrometry (S1 Text for details). The mass spectrometry analyses confirm alterations in lipid A structure of the *ramA* overexpresser, Ecl8Δ*ramR* compared to the wild type (Ecl8), the null mutant (Ecl8Δ*ramA*) or the double mutant (Ecl8Δ*ramRA*) (Fig. 5C) where peaks (m/z 1840, 1866 and 2079) were found to be elevated. Previous studies in *K. pneumoniae* [31, 32] indicate that those peaks correspond to LpxO hydroxylated lipid A species containing a hydroxymyristate group at position 2' as secondary acyl substitution. Therefore, we surmise that RamA

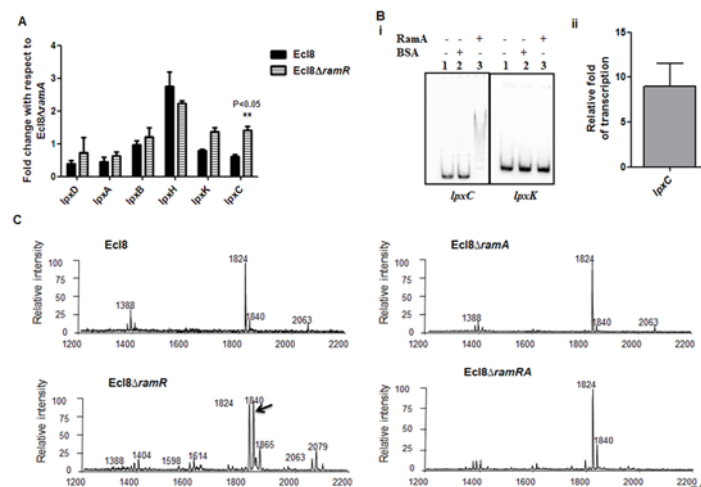


Figure 5. A. Gene Expression analyses of *lpx* genes. All qPCR experiments were performed as outlined in [materials and methods](#). Expression levels were normalized to 16S levels, and fold change values were generated by calibrating against Ecl8ΔramA. Two-way ANOVA analyses ($P < 0.05$) were performed to demonstrate statistical significance. **B. Regulation of *lpx* genes.** (i) EMSA using *lpxC* and *lpxK* promoter regions. Purified RamA (200nM) and the different labelled DNA probes (2 nM) were incubated on ice. All reactions were performed on ice prior to electrophoresis on 7.5% native gel. (ii) Transcription *in vitro* of *lpxC* promoter region. Relative fold increase was determined using densitometric analysis as described previously [55]. Fold increases in the presence of RamA were determined by first normalising to the control promoter (*gnd*) prior to comparison to the no protein control. **C:** Lipid A analysis from *K. pneumoniae* Ecl8 (WT), Ecl8ΔramA, Ecl8ΔramR and Ecl8ΔramRA. Lipid A analysis was undertaken as described before [31]. Negative ion MALDI-TOF mass spectrometry of lipid A isolated from *K. pneumoniae* Ecl8 and its derivatives. Peaks in bold correspond to LpxO dependent 2' secondary chain modifications.

doi:10.1371/journal.ppat.1004627.g005

mediated activation of the different lipid A biosynthetic genes leads to alterations within the lipid A moiety in *K. pneumoniae*.

Antibiotic Susceptibility

Phenotype microarray analyses. In order to assign phenotypes linked to the differentially regulated genes, Biolog phenotype assays were undertaken for *K. pneumoniae* Ecl8 and its isogenic derivatives Ecl8ΔramA and Ecl8ΔramR. A comparison of Biolog phenotypic profiles of both *Salmonella* [11] and *K. pneumoniae* generally indicates a significant overlap in the susceptibilities to antimicrobial and toxic compounds (S3 Table). As expected, the overexpression of *ramA* resulted in increased tolerance of Ecl8ΔramR in the presence of antimicrobials such as tetracyclines (doxycycline, chlortetracycline, minocycline), macrolides (erythromycin, spiramycin, troleandomycin), beta-lactams (1st, 2nd, 3rd generation cephalosporins, penams) and (fluoro)quinolones (ciprofloxacin, ofloxacin, nalidixic acid, novobiocin), fungicides (such as chloroxylonol, dodine, domiphen bromide) and toxic anions (potassium tellurite, sodium metasilicate) (S3 Table, S6 Fig.). Notably, comparisons of the Biolog data also indicate that *ramA* overexpression results in altered polymyxin B susceptibility levels in both *K. pneumoniae* and *Salmonella*.

Susceptibilities to the Polymyxins and the Cationic Antimicrobial Peptides (cAMPs)

Lipid A synthesis in Gram-negative bacteria is controlled at both the transcriptional and translational levels, where alterations in the lipid A profile can result in perturbations in

host-microbe interactions as well as reductions in susceptibility to both the polymyxins and the cationic antimicrobial peptides (cAMPs) [33]. Accordingly, we tested the strain Ecl8 and its isogenic derivatives Ecl8 Δ ramA, Ecl8 Δ ramR against colistin, polymyxin B and the cAMP LL-37. The relative survival assays for colistin, polymyxin B and LL-37 demonstrated that the ramA overexpressing strain, Ecl8 Δ ramR strain was significantly ($P < 0.05$) less susceptible to polymyxin B, colistin and LL-37 (Fig. 6 A, B, C) compared to the wild type Ecl8 and the null mutant Ecl8 Δ ramA. The reduction in polymyxin susceptibility, as noted in the survival assays, is also supported by the Biolog data (S3 Table). Taken together these results suggest that RamA-dependent regulation provides an alternative pathway for reduced susceptibility to polymyxins and cAMPs.

Effect of RamA Overexpression on Host-Microbe Interactions

Macrophage-Klebsiella interaction. To ascertain whether RamA-mediated alterations can have an impact on microbe-macrophage interactions, we examined if Ecl8 and its isogenic derivatives, Ecl8 Δ ramR, Ecl8 Δ ramA and Ecl8 Δ ramRA would exhibit differential interactions in adherence and intracellularization into murine RAW macrophages. In the adhesion and intracellularization assays, the ramA overexpresser, Ecl8 Δ ramR, was significantly attenuated in its ability (approximately 50% decrease) to attach to and internalise into the RAW murine macrophage cells compared to wild type *K. pneumoniae* Ecl8, the mutants Ecl8 Δ ramA and Ecl8 Δ ramRA (Figs. 7A, B and C). Two possible explanations exist for the reduction in adherence and intracellularization of Ecl8 Δ ramR; the first, where altered RamA levels confers resistance to phagocytosis and the second, is due to accelerated killing by the macrophage. In order to ascertain whether the reduced intracellularization of Ecl8 Δ ramR was linked to accelerated killing by macrophages, we determined the levels of extracellular non-phagocytosed bacteria in our experiments and

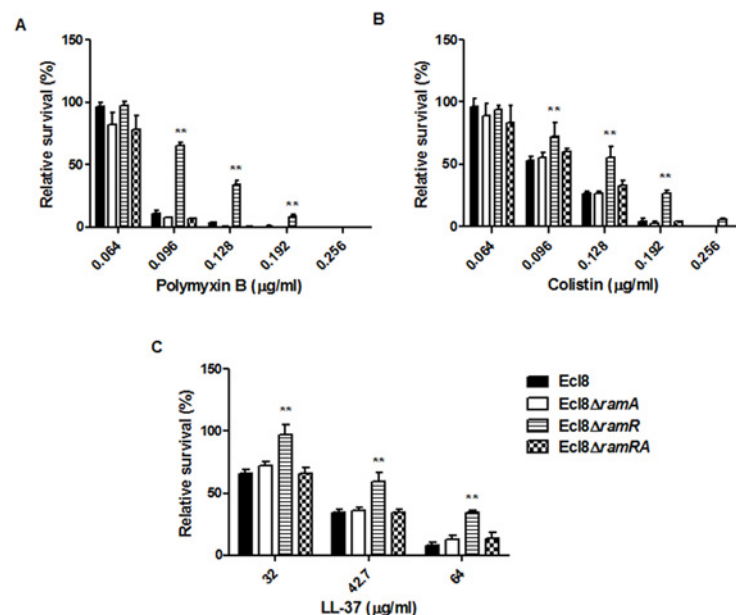


Figure 6. Survival assay of *K. pneumoniae* (Ecl8, Ecl8 Δ ramA, Ecl8 Δ ramR, Ecl8 Δ ramRA) to polymyxin B, colistin and the antimicrobial peptide LL-37. The relative survival of bacteria (expressed as a percentage of the number of colonies obtained from the unexposed control of the same strain) in the presence of different concentrations of polymyxin B (A), colistin (B) and LL-37 (C) are shown. Asterisks indicate that results obtained for the ramA expresser, Ecl8 Δ ramR is significantly different ($P < 0.05$ by Two-way ANOVA) compared to Ecl8, Ecl8 Δ ramA and Ecl8 Δ ramRA.

doi:10.1371/journal.ppat.1004627.g006

found significantly higher numbers of recovered bacteria for Ecl8 Δ ramR compared to the wild type Ecl8, Ecl8 Δ ramA and Ecl8 Δ ramRA (Fig. 7D). In previous work [34], resistance to phagocytosis by *K. pneumoniae* has been linked to bacterial surface structures which include the capsular polysaccharide (cps). However, *ugd* gene transcription, representative of the *cps* cluster [35], was not found to be altered in Ecl8, Ecl8 Δ ramA, Ecl8 Δ ramR and Ecl8 Δ ramRA (S5B Fig.), consistent with the RNAseq data. Thus our results underscore that reduced phagocyte adhesion and uptake is linked to RamA-mediated alterations, particularly those associated with lipid A.

Infection In Vivo

In order to assign a broader relevance to altered *Klebsiella*-host interaction, we performed experiments to assess bacterial recovery using the intranasal inoculation method [36] as described previously. Following a 24-hour infection of 5–7 week old C57BL mice, organ homogenates (spleen and lung) were plated to determine bacterial counts. At 24 h post infection, bacterial recovery rates for the *ramA* overexpressor, Ecl8 Δ ramR were found to be significantly higher compared to the wild type Ecl8 or null mutant Ecl8 Δ ramA from the lung and spleen (Fig. 8(A) and 8(B)). The intranasal route of infection is expected to result in the primary infection of the lung prior to dissemination to other organs. Our results demonstrate that significantly higher levels of Ecl8 Δ ramR is recovered from both the lung and spleen highlighting that RamR-dependent RamA overexpression, confers reduced microbial clearance and increased systemic dissemination of *K. pneumoniae* in an intranasal infection model.

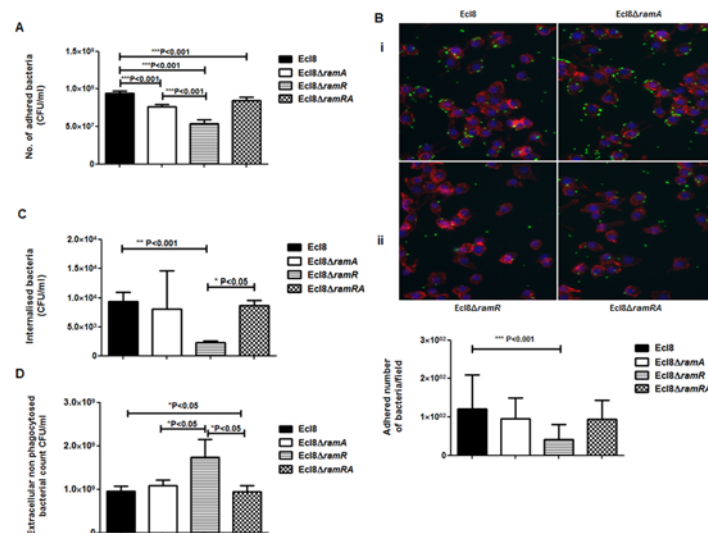


Figure 7. A: Attachment of *K. pneumoniae* Ecl8, Ecl8 Δ ramA, Ecl8 Δ ramR or Ecl8 Δ ramRA to murine macrophage RAW 264.7 cell line. One-way ANOVA analyses were performed to demonstrate statistical significance. **B: Microscopy to assess attachment to RAW 264.7 cell line.** (i) Infection of the RAW264.7 cell line was carried out with *K. pneumoniae* Ecl8 (WT), Ecl8 Δ ramA, Ecl8 Δ ramR or Ecl8 Δ ramRA transformed with plasmid pRSMgfp. MOI was 1:100 and infections were carried out for 2 hrs. The actin cytoskeleton was stained with Acti stain 555 phalloidin (red) and host cell nuclei were stained with DAPI (blue). Images are representative of 80 fields. (ii) Graph representing mean values are derived from 3 independent experiments. One-way ANOVA analyses ($P < 0.001$) were performed to demonstrate statistical significance. **C: Internalisation of *K. pneumoniae* Ecl8, Ecl8 Δ ramA, Ecl8 Δ ramR or Ecl8 Δ ramRA by RAW 264.7 cells.** Bacterial internalisation was assessed by the gentamicin protection assay. One-way ANOVA analyses were performed to demonstrate statistical significance. **D: Enumeration of the extracellular non-phagocytosed *K. pneumoniae* Ecl8, Ecl8 Δ ramA, Ecl8 Δ ramR or Ecl8 Δ ramRA.** One-way ANOVA analyses were performed to demonstrate statistical significance.

doi:10.1371/journal.ppat.1004627.g007

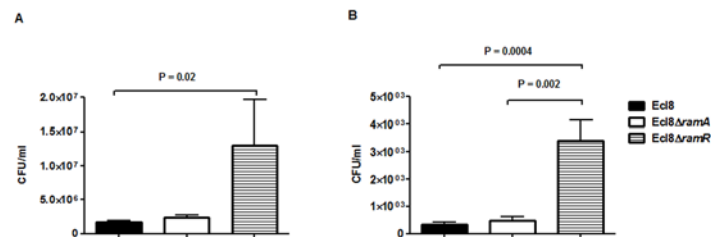


Figure 8. Effect of RamA on bacterial recovery using an intranasal infection model. Bacterial recovery (cfu/ml) was determined from lung (A) and spleen (B) homogenates following a 24h infection of 5–7 week old, C57BL6, female mice (n = 5) using the previously described intranasal infection model. Unpaired t-test analyses were performed to demonstrate statistical significance.

doi:10.1371/journal.ppat.1004627.g008

Discussion

The relevance of the MarA, SoxS, Rob, RamA and RarA regulators in microbial survival is attributed to their control of the antimicrobial resistance phenotype in a wide variety of Gram-negative bacteria [10, 37, 38]. Whilst the role of RamA in reduced antibiotic susceptibility is evident from multiple studies [16, 17, 37], its broader role in gene regulation is not known in *Klebsiella pneumoniae*. Using transcriptome profiling, we demonstrate that RamA-overexpression results in altered *K. pneumoniae* transcription patterns (S3 Fig.) compared to the null mutant or wild type strain thus highlighting its wider role in gene regulation in *K. pneumoniae*.

Our data suggests that RamA functions largely as a transcriptional activator of gene expression, where DNA-binding (Fig. 4B) and IVT assays (Fig. 4C) demonstrate that this regulation is direct and likely mediated via a *mar/ram*-box like element [39] located within the promoter region. Whilst our work is the first to demonstrate direct RamA-mediated activation of gene expression, other studies have shown that related proteins such as MarA, SoxS [40] and RarA [5] also exert explicit control of regulon genes. Comparative transcriptome data analyses suggests that RamA-mediated activation is dependent on regulator concentration (basal versus overexpressed, Fig. 2) in addition to the observation that identical RamA levels induce differential levels of promoter activation as supported by our *in vitro* data (Fig. 4C). The maintenance of basal *ramA* levels may be necessary for the *K. pneumoniae* stress response to a variety of agents as has been previously shown when selecting for fluoroquinolone resistant *Salmonella* [41] or *Klebsiella* in a *ramA*-deleted strain. In *K. pneumoniae*, basal levels of *ramA* expression is maintained due to titration of the absolute repressory effects of RamR by the RamR-sRamA5 interaction (S2 Fig.). Uniquely for tetracycline family regulators, RamR, directly interacts with the regulatory RNA, sRamA5, (Fig. 2B) which is produced as a cleaved by-product of the primary *romA* transcript (Fig. 2B). Whilst the sRamA5-RamR interaction, provides basal levels of *ramA* expression, *ramA* transcription as observed in the overexpressor, Ecl8ΔramR or clinical strains [16] are linked to loss of function mutations within RamR. Consequently, our data show that the maximal changes in gene expression profiles are observed when *ramA* is overexpressed as in Ecl8ΔramR (S3 Fig.). In this gene cohort, we demonstrate that RamA impacts on gene transcription linked to operons associated with efflux pumps, biofilm formation and lipid A biosynthesis (Fig. 3, S2 Table). Whilst it is possible that the differential regulation of these genes is not all directly linked to RamA, we demonstrate that purified RamA directly binds and activates the expression of multiple associated promoters (Fig. 4C & 4D).

A comparison of RamA-mediated regulation in *Salmonella enterica* serovar Typhimurium [4] and *K. pneumoniae* establishes key similarities in the genes associated with the respective RamA regulons; particularly in the control of genes associated with antimicrobial resistance

acrAB and *ompF* [4, 11]. Additionally, RamA-dependent direct activation of *acrAB* is also consistent with phenotypic studies [10, 16–18] which consistently demonstrate that *ramA* overexpression is linked to increased elevated efflux via *acrAB* and decreased outer membrane protein levels (OmpF). Given its role in conferring reduced antimicrobial susceptibility, it is perhaps not surprising that we demonstrate that RamA directly regulates other efflux related operons specifically; the AcrAB linked inner periplasmic protein, YbhT [42] associated with detergent sensitivity, the Yrb operon which encodes an ABC transporter linked to the export of quinolones [27] and also lipid asymmetry [30]. The combined effect of the efflux or influx levels and membrane alterations associated with transport and structural variations likely contributes to the substrate range of compounds impacted by *ramA* overexpression (S3 Table). However, in the absence of a functional *acrAB* efflux pump, RamA-overexpression does not confer reduced susceptibility to most antibiotics in *K. pneumoniae*. This observation is consistent with previous studies for the MarA and RarA [38] proteins.

Therefore, it is likely that a functional AcrAB pump is crucial in mediating decreased antimicrobial susceptibility. However, a recent study [43] also suggests that *acrAB* may play a role in decreased antimicrobial peptide susceptibility and increased virulence in *K. pneumoniae*. Our findings support this observation and further demonstrate that increased RamA levels can also mediate LPS alterations, which likely contribute towards increased survival to both polymyxins and cationic AMPs (Fig. 5, 6).

Structurally, LPS is composed of three domains, the serovar dependent O-antigen chain, core oligosaccharide consisting of sugars and lipid A which is a phosphorylated disaccharide decorated with multiple fatty acids which anchor the LPS into the bacterial membrane [29]. The endotoxic lipid A component of LPS constitutes the outermost layer of the outer membrane of Gram-negative bacteria thereby playing a critical role in host-microbe interactions in addition to promoting reduced susceptibility to cAMPs [44] such as polymyxins [30] and host derived factors LL-37, HBD-1 [30]. Studies have shown that lipid A modifications can result in multiple outcomes such as reduced polymyxin susceptibility [45] in addition to directly facilitating microbial evasion by reduced immune recognition [46]. Our work suggests that the molecular basis for the modified lipid A structure is linked to the differential regulation of the biosynthesis genes e.g. *lpxO*, *lpxL-2* and *lpxC* identified in this screen. Despite being constitutively produced the regulation of *lpxC*, *lpxL-2* and *lpxO*, is still subject to either transcriptional or translational control [44, 46]; generally in response to stress, where, *lpxC* and *lpxL-2* are regulated by the two-component systems, PhoPQ and PmrAB [44]. In contrast, *lpxO* is not subject to PhoPQ regulation in *Salmonella* [44, 46].

In *Salmonella* Typhimurium, the modulation of LpxO levels results in the remodeling of the outer membrane which reduces the net negative charge whilst simultaneously increasing membrane integrity resulting in increased virulence [47]. A similar phenotype is exhibited by the *K. pneumoniae* Ecl8Δ*ramR* strain, which has altered LpxO levels (Fig. 8). Thus we surmise that the altered host-microbe and polymyxin-microbe interactions are in part attributable to the lipid A modifications.

Macrophages represent a key innate host defence strategy against microbial infections as phagocytosis of incoming pathogens is a trigger for the inflammatory response. Our data show that *ramA* overexpression protects against macrophage uptake and internalization (Fig. 7) thus providing a basis for the greater dissemination of the *ramA* overexpressing strain, Ecl8Δ*ramR* in an *in vivo* infection model. Taken together, these RamA-linked phenotypes underscore its' role in *Klebsiella* virulence and survival *in vivo*.

The molecular basis for phenotypes associated with reduced antimicrobial peptide susceptibility and increased virulence can be attributed to several key loci such as the *acrAB* pump and lipid A biosynthesis genes, *lpxC*, *lpxL-2* and *lpxO*. This is supported by studies that

demonstrate the involvement of *acrAB* [48] and lipid A modifications [30, 44] in host-microbe interactions. However to definitively pinpoint the exact contribution of the lipid A biosynthetic genes or *acrAB* to phenotypes associated with host-pathogen interactions would require the deletion of genes encoding *lpxC* [49], *lpxL-2* and *lpxO* [50], *acrAB* individually or in combination with *ramA* overexpression. We note that previous studies [32, 50] have shown that strains deleted for these genes, result in avirulent microbes and as such, this phenotype would obscure any RamA-associated effects. Nevertheless, our work is first to demonstrate that firstly, RamA functions as an alternate regulator of certain lipid A biosynthesis genes and secondly, these alterations perturb microbe-host interaction.

The significance of our findings lies in the broader implications of RamA-mediated regulation in Enterobacteriaceae. In this work, we describe roles for RamA in both protection against antibiotic challenge but also against the innate host immune response thus resulting in *Klebsiellae* that are less susceptible to antibiotics and simultaneously more virulent. Notably, our findings highlight that RamA mediated overexpression via both increased *acrAB* expression and lipid A alterations can result in reduced susceptibility to the last line drugs e.g. tigecycline and polymyxins. This highlights the broader consequences in selecting for *ramA* overexpression in *K. pneumoniae* or other members of Enterobacteriaceae. Finally, our study underscores and highlights the importance of intrinsic proteins such as RamA, which regulate survival strategies in *K. pneumoniae* and likely other Enterobacteriaceae, specifically in priming the microbial population in surviving drug and host immune pressure. This proposes the notion where microbial immune evasive strategies contribute to the development and persistence of antimicrobial resistance.

Materials and Methods

Growth Conditions

Bacteria (Table 1) were cultured in Luria-Bertani (LB) medium (10 g/L tryptone, 5 g/L yeast extract, 10 g/L NaCl). Typically, a strain was first grown on an LB plate at 37°C from frozen -80°C stocks. A single colony was picked and inoculated into 5 ml of LB and incubated in a 37°C shaker overnight. A 1 in 100 dilution was made in LB and incubated in a 37°C shaker until the OD₆₀₀ reached 0.6 unless otherwise stated. Antibiotics such as ampicillin (100 µg/ml) and chloramphenicol (20 µg/ml) were added as required.

Antimicrobial Susceptibility and Peptide Survival Assay

The assay was as described previously by Moranta *et al* [51]. Briefly, bacteria were grown at 37°C in 5 ml LB medium, harvested (5,000 × g, 15 min, 5°C) and washed thrice with phosphate-buffered saline (PBS). A suspension containing approximately 10⁵ CFU/ml was prepared in 10 mM PBS (pH 6.5), 1% tryptone soy broth (TSB; Oxoid), and 100 mM NaCl. Aliquots (5 µl) of this suspension were mixed in tubes with various concentrations of polymyxin B, colistin (0.064 µg/ml to 0.256 µg/ml) and LL-37 (32 µg/ml to 85.3 µg/ml) to a final volume of 30 µl. Following incubation for an hour at 37°C with polymyxin B (Sigma, UK), colistin

Table 1. Strains used in this study.

Strains	Description	Ref
Ecl8	Wild type <i>K. pneumoniae</i>	[59]
Ecl8Δ <i>ramA</i>	<i>K. pneumoniae</i> deleted of <i>ramA</i>	[16]
Ecl8Δ <i>ramR</i>	<i>K. pneumoniae</i> deleted of <i>ramR</i>	[16]

doi:10.1371/journal.ppat.1004627.t001

(Sigma, UK) and LL-37 (Sigma, UK) the samples were diluted 1:10 with PBS prior to plating (100 μ l) on LB agar. Colony counts were determined after overnight incubation, where results are expressed as percentages of the colony count of bacteria that were not exposed to the antibiotics or the antimicrobial peptide. Sensitivity profiles of the different mutants using the phenotypic microarray analyses were determined described in [S1 Text](#).

RNA Extraction, RNA-Seq Sample Preparation and Sequencing

Overnight cultures of strains Ecl8, Ecl8 Δ ramA, Ecl8 Δ ramR were inoculated (1/100 dilution) into LB media and incubated at 37 °C with vigorous shaking. Cell pellets were harvested at OD₆₀₀ = 0.6 and RNA was extracted using the RNAeasy Extraction Kit (Qiagen, Hilden, Germany), which enriches for RNA molecules larger than 200 nucleotides. No depletion of ribosomal RNA was carried out prior to the synthesis of single-stranded cDNA (sscDNA) as previously reported [52]. RNAseq DNA libraries were constructed as previously described [53]. For RNA-seq, independent biological samples in triplicate were assessed for each strain. The resulting sscDNA libraries were sequenced in an Illumina HiSeq 2000 sequencer. An average of 0.715 Gb of sequence data was obtained per sample, in 75 bp paired reads (Details of RNAseq analyses are outlined in [S1 Text](#)). The RNAseq read data has been deposited under the ENA data repository and ArrayExpress with the accession numbers ERP001994 and E-ERAD-122, respectively.

RNA for quantitative Real-Time PCR experiments was extracted from *K. pneumoniae* strains ([Table 1](#)) using the TRIzol extraction method [16]. Briefly, cells were grown to mid-log phase (OD₆₀₀ = 0.6) at 37 °C with shaking and then harvested by centrifugation at 3000g (PK121R, ALC) at 4 °C. The cell pellet was then resuspended in TRIzol reagent (Invitrogen, Paisley, UK) and chloroform prior to centrifugation to separate the phases. The upper phase was then precipitated using 3 M sodium acetate, glycogen (5 mg/ml), and 100% ethanol.

Both RNA preparations were washed and resuspended in 50 μ l DEPC treated water. RNA was treated with TurboDNase to remove DNA contamination (Ambion, New York, USA). All samples were assessed for RNA integrity and quantification using both the Bioanalyzer 2100 RNA nanochip (Agilent, UK) and the ND-1000 (Nanodrop Technologies) [4]. Only those samples with integrity level 9 were taken forward for library construction or qPCR analyses.

Quantitative Real-Time PCR (qPCR)

In order to validate the RNAseq data, quantitative Real-Time PCR experiments were undertaken. After the removal of contaminating DNA, cDNA synthesis was generated using the AffinityScript cDNA synthesis kit (Agilent, UK). Gene specific primers ([S4 Table](#)) were designed using the Primer3 (<http://frodo.wi.mit.edu/>) software and were tested to produce standard curves with amplification efficiencies ranging from 95–110%. qPCR analyses using the locked nucleic acid probe is detailed in [S1 Text](#). Quantitative Real Time RT-PCR (RT-PCR) was performed using the synthesized cDNA with gene specific primers using the Brilliant III Ultra-fast SYBR Green Kit (Agilent, UK) in the Agilent Mx3005P. All data were analyzed using Agilent MxPro software, which is based on the efficiency corrected method (Pfaffl) of comparative quantification that utilizes the $\Delta\Delta$ Ct approach, also taking into account primer efficiency. The relative fold increases in expression levels were determined by firstly normalizing gene expression levels to 16S rDNA and using either Ecl8 or Ecl8 Δ ramA as calibrators. All comparative analyses were done using the MxPro software (Agilent, UK).

DNA EMSA

DNA fragments that represent the promoter regions of the genes that were differentially regulated in the presence or absence of RamA or RamR were subjected to the electrophoretic gel

shift mobility assay (EMSA) as described previously [54]. Briefly, DNA templates ranging from 250–150bp upstream of the start site were produced by PCR, and purified by StrataPrep PCR Purification kits (Agilent UK). The purified templates were end-labelled with [$\gamma^{32}\text{P}$]-ATP by T4 Kinase (New England Biolabs, USA). The unincorporated, labelled ATP was removed using Biospin P6 spin columns (Biorad, UK) as per manufacturer's instructions. Purified RamA was extracted from the recombinant pETramA construct using metal chelation chromatography on superflow nickel / nitrilotriacetate agarose (Qiagen, Germany) (James Hastie, Dundee University). His-tagged RamA (200 nM) and labelled DNA (2 nM) were mixed in binding buffer (125 mM Tris-HCl, 250 mM KCl, 5 mM DTT 5% glycerol) and incubated on ice for 15 min prior to electrophoresis at 75 V on a prechilled 7.5% native polyacrylamide gel in $1 \times$ TBE buffer.

Transcription *In Vitro*

Transcription (IVT) experiments were performed as described previously [55]. Briefly $5 \times$ IVT Buffer, 2 nM PCR product of the test and control (*E. coli gnd* [56]) promoters, RNA polymerase, RNaseOUT (Invitrogen, UK) was incubated for 15 minutes at 37°C prior to the addition of the transcription mix containing $\times 5$ IVT buffer (50 mM Tris-HCl, 0.1 mM EDTA, 3 mM magnesium acetate, 0.1 mM dithiothreitol, 20 mM sodium chloride, and 250 $\mu\text{g}/\text{ml}$ bovine serum albumin at pH 7.8), heparin (1.2 $\mu\text{g}/\text{ml}$), NTPS, and $\alpha^{32}\text{P}$ -UTP (Perkin Elmer, UK). The reaction was stopped 5 minutes later followed by the addition of Gel loading buffer II (Ambion, UK). The resulting products were electrophoresed on a 7% polyacrylamide / 8 M urea gel. Quantification was determined by densitometric analysis using Fujifilm Multigauge Software where an increase or decrease in transcription levels is after normalization to the endogenous *gnd* levels and calibration to the no protein control.

Cell Culture

Murine RAW 264.7 macrophage cells (obtained from ATCC TIB-71) were cultured in Dulbecco's Modified Eagle Medium (PAA, UK) supplemented with 10% endotoxin-free foetal bovine serum (PAA, UK) and penicillin and streptomycin (Invitrogen, UK) in 75- cm^2 culture flasks in 5% CO_2 for 24 h until subconfluent. Twelve well tissue culture plates were seeded with 5×10^5 cells per well and viability determined using trypan blue exclusion.

Bacterial Adhesion and Internalisation Assays

Bacterial adhesion and internalization experiments were performed as described previously [57, 58]. For the adhesion assays, RAW cells were washed with PBS and incubated for 2 h at 37°C in 5% CO_2 with a suspension of 5×10^7 bacterial cells in DMEM medium alone. After incubation, wells were washed five times with PBS and adherent bacteria were released by addition of 0.5% Triton X-100 (Sigma, UK). Bacterial colonies were quantified by plating appropriate dilutions on LB agar plates. In the internalization assays, after the incubation of the RAW cells with the bacterial suspension, wells were washed twice with PBS and then incubated for 2 h with fresh DMEM medium containing gentamicin (100 $\mu\text{g}/\text{ml}$) to eliminate extracellular bacteria. After the incubation, an aliquot of the medium was plated to confirm killing of extracellular bacteria and the gentamicin-containing medium was washed again. RAW cells were lysed and intracellular bacteria were quantified as described above. To estimate levels of extracellular bacteria, the infection of the RAW cells was carried out as described previously for the adhesion assay. After incubation, the media with the non-phagocytosed extracellular bacteria was collected and quantified by plating appropriate dilutions on LB agar plates. All microscopy images were generated as outlined in [S1 Text](#).

Ethics Statement

All mouse experiments were performed under the control of the UK Home Office legislation in accordance with the terms of the Project license (PPL2700) granted for this work under the Animals (Scientific Procedures) Act 1986 in addition to receiving formal approval of the document through Queen's University Belfast Animal Welfare and Ethical Review Body.

Infection *In Vivo*

Overnight bacterial cultures were washed three times in sterile endotoxin free PBS. The bacteria was resuspended to an optical density of 0.2 and 20 μ l ($\sim 5 \times 10^7$ CFU/animal) and administered to anaesthetised 5–7 week old weight watched Harlan C57BL6 mice ($n = 5$ per group) using the intranasal inoculation method [36]. In order to ensure maximal delivery of the bacterial inoculation into the lungs the animals were held in a perpendicular position until cessation of laboured breathing. 24 h post inoculation the mice were sacrificed by lethal pentobarbitol injection. Perfused lungs and spleen were harvested and resuspended in 1 ml of sterile PBS. Following mechanical homogenisation dilutions were plated on LB agar plates and incubated at 37°C to establish the CFU/ml.

Supporting Information

S1 Fig. Transcriptional start sites (TSS) of *ramR*, *sRamA5* and *romA* determined by 5' RACE. Capitalized triplets are start codons of either *ramR* or *romA*. Bold capital letters indicate the primary and secondary TSS sites for *ramR* or *sRamA5* and *romA*. The primary and secondary TSSs of *ramR* are indicated with a black and grey arrow respectively. The shaded segments are the inverted repeat (IR) sequences recognized by the RamR protein. The sequences indicated by the dotted underlined fragment (RL) and the single underlined fragment (RH) were used in eGFP analysis. The numbering system is based on the "t" prior to *romA*'s start codon ATG as the -1. (TIF)

S2 Fig. Fluorescence reporter analysis for RH promoter activity and its repression by RamR. Where indicated DH5 α contained both the pKC026 (containing either the RH fragments) and pBR322*ramR* plasmid. DH5 α harboring both pKC026 and pBR322 is taken as a negative control and the RH fragment containing pKC026 with pBR322 only taken as the baseline control. (TIF)

S3 Fig. Kolmogorov-Smirnov 2-sample test. The plots show the sample distribution of log₂-transformed basemean values from the RNAseq experiment for Ecl8, Ecl8 Δ *ramA* and Ecl8 Δ *ramR*. The D value (the largest vertical distance between two curves) between Ecl8 and Ecl8 Δ *ramA* is 0.046; the one between Ecl8 and Ecl8 Δ *ramR* is 0.067, and the one between Ecl8 Δ *ramA* and Ecl8 Δ *ramR* is 0.036. All of the distributions are significantly different from each other ($p < 0.001$). (TIF)

S4 Fig. Coverage plot for the antisense transcription of BN373_16241 in Ecl8 Δ *ramA* and Ecl8 Δ *ramR*. The coverage plot is visualized with Artemis Genome Browser using the Ecl8 genome as a reference. Window size is set at 3. One representative coverage plot of Ecl8 Δ *ramA* and Ecl8 Δ *ramR* is shown here. The borders of the coding region of BN373_16241 are marked by vertical black bars in the coverage plot. The green and red curves within the borders represent the antisense and sense transcription of BN373_16241 respectively. (TIF)

S5 Fig. A: Quantitative real-time RT-PCR analyses of *romA* and *ramA* gene expression levels in Ecl8Δ*ramR*. B: Gene expression of *ugd*, *pagP* genes in different *K. pneumoniae* strains. All qPCR experiments were performed as outlined in [materials and methods](#). Expression levels were normalized to 16S levels, and fold change values were generated by calibrating against Ecl8Δ*ramA*. All data is a mean of 3 experiments.

(TIF)

S6 Fig. Heat map of the Biolog phenotype of Ecl8, Ecl8Δ*ramA* and Ecl8Δ*ramR*. Biolog analyses of the wild type *K. pneumoniae* Ecl8, Ecl8Δ*ramA* and *K. pneumoniae* Ecl8Δ*ramR* using PM1–20 plates.

(TIF)

S1 Text. Supporting Materials and Methods.

(DOCX)

S1 Table. Raw mapped RNAseq data.

(XLS)

S2 Table. List of differentially expressed genes in the pairwise comparison with COG analyses.

(XLSX)

S3 Table. Biolog phenotypic profile of the *K. pneumoniae* stains.

(XLSX)

S4 Table. List of primers used in this study.

(XLSX)

Acknowledgments

The authors acknowledge reagent, technical support and advice provided by Nuala Tipping, Tom Gardiner, Eimear Linehan and Denise Fitzgerald, Centre for Infection and Immunity, QUB, for microscopy reagents. We thank Dr. Nicola Holden, Prof. Jay Hinton for the plasmid pRSMgfp+, Dr. Bhargav Mitra for the program code in Matlab for microscopy data analyses and Prof. N. Thomson for facilitating the transcriptome analyses.

Author Contributions

Conceived and designed the experiments: SDM JY TS. Performed the experiments: SDM JY MF SPM EL SFi AM. Analyzed the data: SDM JY MF SFi RJI JB TS. Contributed reagents/materials/analysis tools: SDM JY SPM MF EL SFi SS AM AK RJI JB DLG SFa JSE TS. Wrote the paper: SDM JY MF TS.

References

1. Barbosa TM, Levy SB (2000) Differential Expression of Over 60 Chromosomal Genes in *Escherichia coli* by Constitutive Expression of MarA. *J Bacteriol* 182: 3467–3474. doi: [10.1128/JB.182.12.3467-3474.2000](#) PMID: [10852879](#)
2. Pomposiello PJ, Bennik MH, Demple B (2001) Genome-wide Transcriptional Profiling of the *Escherichia coli* Responses to Superoxide Stress and Sodium Salicylate. *J Bacteriol* 183: 3890–3902. doi: [10.1128/JB.183.13.3890-3902.2001](#) PMID: [11395452](#)
3. Bennik MH, Pomposiello PJ, Thorne DF, Demple B (2000) Defining a rob regulon in *Escherichia coli* by using transposon mutagenesis. *J Bacteriol* 182: 3794–3801. doi: [10.1128/JB.182.13.3794-3801.2000](#) PMID: [10850996](#)
4. Bailey AM, Ivens A, Kingsley R, Cottell JL, Wain J, et al. (2010) RamA, a member of the AraC/XylS family, influences both virulence and efflux in *Salmonella enterica* serovar Typhimurium. *J Bacteriol* 192: 1607–1616. doi: [10.1128/JB.01517-09](#) PMID: [20081028](#)

5. De Majumdar S, Veleba M, Finn S, Fanning S, Schneiders T (2013) Elucidating the regulon of multidrug resistance regulator RarA in *Klebsiella pneumoniae*. *Antimicrob Agents Chemother* 57: 1603–1609. doi: [10.1128/AAC.01998-12](https://doi.org/10.1128/AAC.01998-12) PMID: [23318802](https://pubmed.ncbi.nlm.nih.gov/23318802/)
6. Martin RG, Rosner JL (2001) The AraC transcriptional activators. *Curr Opin Microbiol* 4: 132–137. doi: [10.1016/S1369-5274\(00\)00178-8](https://doi.org/10.1016/S1369-5274(00)00178-8) PMID: [11282467](https://pubmed.ncbi.nlm.nih.gov/11282467/)
7. Piddock LJ (2006) Multidrug-resistance efflux pumps—not just for resistance. *Nat Rev Microbiol* 4: 629–636. doi: [10.1038/nrmicro1464](https://doi.org/10.1038/nrmicro1464) PMID: [16845433](https://pubmed.ncbi.nlm.nih.gov/16845433/)
8. Dangi B, Pelupessey P, Martin RG, Rosner JL, Louis JM, et al. (2001) Structure and Dynamics of MarA-DNA Complexes: An NMR Investigation. *J Mol Biol* 314: 113–127. doi: [10.1006/jmbi.2001.5106](https://doi.org/10.1006/jmbi.2001.5106) PMID: [11724537](https://pubmed.ncbi.nlm.nih.gov/11724537/)
9. Griffith KL, Shah IM, Myers TE, O'Neill MC, Wolf RE Jr. (2002) Evidence for “pre-recruitment” as a new mechanism of transcription activation in *Escherichia coli*: the large excess of SoxS binding sites per cell relative to the number of SoxS molecules per cell. *Biochem Biophys Res Commun* 291: 979–986. doi: [10.1006/bbrc.2002.6559](https://doi.org/10.1006/bbrc.2002.6559) PMID: [11866462](https://pubmed.ncbi.nlm.nih.gov/11866462/)
10. Alekshun MN, Levy SB (1997) Regulation of Chromosomally Mediated Multiple Antibiotic Resistance: the *mar* Regulon. *Antimicrob Agents Chemother* 41: 2067–2075. PMID: [9333027](https://pubmed.ncbi.nlm.nih.gov/9333027/)
11. Zheng J, Tian F, Cui S, Song J, Zhao S, et al. (2011) Differential gene expression by RamA in ciprofloxacin-resistant *Salmonella* Typhimurium. *PLoS One* 6: e22161. doi: [10.1371/journal.pone.0022161](https://doi.org/10.1371/journal.pone.0022161) PMID: [21811569](https://pubmed.ncbi.nlm.nih.gov/21811569/)
12. Hung DT, Shakhnovich EA, Pierson E, Mekalanos JJ (2005) Small-molecule inhibitor of *Vibrio cholerae* virulence and intestinal colonization. *Science* 310: 670–674. doi: [10.1126/science.1116739](https://doi.org/10.1126/science.1116739) PMID: [16223984](https://pubmed.ncbi.nlm.nih.gov/16223984/)
13. Casaz P, Garrity-Ryan LK, McKenney D, Jackson C, Levy SB, et al. (2006) MarA, SoxS and Rob function as virulence factors in an *Escherichia coli* murine model of ascending pyelonephritis. *Microbiology* 152: 3643–3650. doi: [10.1099/mic.0.2006/000604-0](https://doi.org/10.1099/mic.0.2006/000604-0) PMID: [17159217](https://pubmed.ncbi.nlm.nih.gov/17159217/)
14. Chollet R, Chevalier J, Bollet C, Pages JM, Davin-Regli A (2004) RamA is an alternate activator of the multidrug resistance cascade in *Enterobacter aerogenes*. *Antimicrob Agents Chemother* 48: 2518–2523. doi: [10.1128/AAC.48.7.2518-2523.2004](https://doi.org/10.1128/AAC.48.7.2518-2523.2004) PMID: [15215103](https://pubmed.ncbi.nlm.nih.gov/15215103/)
15. Yassien MA, Ewis HE, Lu CD, Abdelal AT (2002) Molecular cloning and characterization of the *Salmonella enterica* Seroovar Paratyphi B *rma* Gene, which confers multiple drug resistance in *Escherichia coli*. *Antimicrob Agents Chemother* 46: 360–366. doi: [10.1128/AAC.46.2.360-366.2002](https://doi.org/10.1128/AAC.46.2.360-366.2002) PMID: [11796342](https://pubmed.ncbi.nlm.nih.gov/11796342/)
16. Rosenblum R, Khan E, Gonzalez G, Hasan R, Schneiders T (2011) Genetic regulation of the *ramA* locus and its expression in clinical isolates of *Klebsiella pneumoniae*. *Int J Antimicrob Agents* 38: 39–45. doi: [10.1016/j.ijantimicag.2011.02.012](https://doi.org/10.1016/j.ijantimicag.2011.02.012) PMID: [21514798](https://pubmed.ncbi.nlm.nih.gov/21514798/)
17. Hentschke M, Wolters M, Sobottka I, Rohde H, Aepfelbacher M (2010) *ramR* mutations in clinical isolates of *Klebsiella pneumoniae* with reduced susceptibility to Tigecycline. *Antimicrob Agents Chemother* 54: 2720–2723. doi: [10.1128/AAC.00085-10](https://doi.org/10.1128/AAC.00085-10) PMID: [20350947](https://pubmed.ncbi.nlm.nih.gov/20350947/)
18. Abouzeed YM, Baucheron S, Cloeckaert A (2008) *ramR* mutations involved in efflux-mediated multidrug resistance in *Salmonella enterica* serovar Typhimurium. *Antimicrob Agents Chemother* 52: 2428–2434. doi: [10.1128/AAC.00084-08](https://doi.org/10.1128/AAC.00084-08) PMID: [18443112](https://pubmed.ncbi.nlm.nih.gov/18443112/)
19. Yamasaki S, Nikaido E, Nakashima R, Sakurai K, Fujiwara D, et al. (2013) The crystal structure of multidrug-resistance regulator RamR with multiple drugs. *Nat Commun* 4: 2078. doi: [10.1038/ncomms3078](https://doi.org/10.1038/ncomms3078) PMID: [23800819](https://pubmed.ncbi.nlm.nih.gov/23800819/)
20. Martin RG, Bartlett ES, Rosner JL, Wall ME (2008) Activation of the *Escherichia coli* *marA/soxS/rob* regulon in response to transcriptional activator concentration. *J Mol Biol* 380: 278–284. doi: [10.1016/j.jmb.2008.05.015](https://doi.org/10.1016/j.jmb.2008.05.015) PMID: [18514222](https://pubmed.ncbi.nlm.nih.gov/18514222/)
21. Ruzin A, Visalli MA, Keeney D, Bradford PA (2005) Influence of transcriptional activator RamA on expression of multidrug efflux pump AcrAB and tigecycline susceptibility in *Klebsiella pneumoniae*. *Antimicrob Agents Chemother* 49: 1017–1022. doi: [10.1128/AAC.49.3.1017-1022.2005](https://doi.org/10.1128/AAC.49.3.1017-1022.2005) PMID: [15728897](https://pubmed.ncbi.nlm.nih.gov/15728897/)
22. Podschun R, Ullmann U (1998) *Klebsiella* spp. as nosocomial pathogens: epidemiology, taxonomy, typing methods, and pathogenicity factors. *Clin Microbiol Rev* 11: 589–603.
23. Fookes M, Yu J, De Majumdar S, Thomson N, Schneiders T (2013) Genome Sequence of *Klebsiella pneumoniae* Ecl8, a Reference Strain for Targeted Genetic Manipulation. *Genome Announc* 1. doi: [10.1128/genomeA.00027-12](https://doi.org/10.1128/genomeA.00027-12)
24. Chinni SV, Raabe CA, Zakaria R, Randau G, Hoe CH, et al. (2010) Experimental identification and characterization of 97 novel npcRNA candidates in *Salmonella enterica* serovar Typhi. *Nucleic Acids Res*. doi: [10.1093/nar/gkq281](https://doi.org/10.1093/nar/gkq281)

25. Yoder-Himes DR, Chain PS, Zhu Y, Wurtzel O, Rubin EM, et al. (2009) Mapping the *Burkholderia cenocepacia* niche response via high-throughput sequencing. *Proc Natl Acad Sci U S A* 106: 3976–3981. doi: [10.1073/pnas.0813403106](https://doi.org/10.1073/pnas.0813403106) PMID: [19234113](https://pubmed.ncbi.nlm.nih.gov/19234113/)
26. Pomposiello PJ, Demple B (2000) Identification of SoxS-regulated genes in *Salmonella enterica* serovar typhimurium. *J Bacteriol* 182: 23–29. doi: [10.1128/JB.182.1.23-29.2000](https://doi.org/10.1128/JB.182.1.23-29.2000) PMID: [10613858](https://pubmed.ncbi.nlm.nih.gov/10613858/)
27. Han X, Geng J, Zhang L, Lu T (2011) The role of *Escherichia coli* YrbB in the lethal action of quinolones. *J Antimicrob Chemother* 66: 323–331. doi: [10.1093/jac/dkq427](https://doi.org/10.1093/jac/dkq427) PMID: [21098540](https://pubmed.ncbi.nlm.nih.gov/21098540/)
28. Malinverni JC, Silhavy TJ (2009) An ABC transport system that maintains lipid asymmetry in the gram-negative outer membrane. *Proc Natl Acad Sci U S A* 106: 8009–8014. doi: [10.1073/pnas.0903229106](https://doi.org/10.1073/pnas.0903229106) PMID: [19383799](https://pubmed.ncbi.nlm.nih.gov/19383799/)
29. Raetz CR, Guan Z, Ingram BO, Six DA, Song F, et al. (2009) Discovery of new biosynthetic pathways: the lipid A story. *J Lipid Res* 50 Suppl: S103–108. doi: [10.1194/jlr.R800060-JLR200](https://doi.org/10.1194/jlr.R800060-JLR200) PMID: [18974037](https://pubmed.ncbi.nlm.nih.gov/18974037/)
30. Gunn JS (2001) Bacterial modification of LPS and resistance to antimicrobial peptides. *J Endotoxin Res* 7: 57–62. doi: [10.1177/09680519010070011001](https://doi.org/10.1177/09680519010070011001) PMID: [11521084](https://pubmed.ncbi.nlm.nih.gov/11521084/)
31. Llobet E, Campos MA, Gimenez P, Moranta D, Bengoechea JA (2011) Analysis of the networks controlling the antimicrobial-peptide-dependent induction of *Klebsiella pneumoniae* virulence factors. *Infect Immun* 79: 3718–3732. doi: [10.1128/IAI.05226-11](https://doi.org/10.1128/IAI.05226-11) PMID: [21708987](https://pubmed.ncbi.nlm.nih.gov/21708987/)
32. Clements A, Tull D, Jenney AW, Farn JL, Kim SH, et al. (2007) Secondary acylation of *Klebsiella pneumoniae* lipopolysaccharide contributes to sensitivity to antibacterial peptides. *J Biol Chem* 282: 15569–15577. doi: [10.1074/jbc.M701454200](https://doi.org/10.1074/jbc.M701454200) PMID: [17371870](https://pubmed.ncbi.nlm.nih.gov/17371870/)
33. Campos MA, Vargas MA, Regueiro V, Llompart CM, Alberti S, et al. (2004) Capsule polysaccharide mediates bacterial resistance to antimicrobial peptides. *Infect Immun* 72: 7107–7114. doi: [10.1128/IAI.72.12.7107-7114.2004](https://doi.org/10.1128/IAI.72.12.7107-7114.2004) PMID: [15557634](https://pubmed.ncbi.nlm.nih.gov/15557634/)
34. March C, Cano V, Moranta D, Llobet E, Perez-Gutierrez C, et al. (2013) Role of bacterial surface structures on the interaction of *Klebsiella pneumoniae* with phagocytes. *PLoS One* 8: e56847. doi: [10.1371/journal.pone.0056847](https://doi.org/10.1371/journal.pone.0056847) PMID: [23457627](https://pubmed.ncbi.nlm.nih.gov/23457627/)
35. Shu HY, Fung CP, Liu YM, Wu KM, Chen YT, et al. (2009) Genetic diversity of capsular polysaccharide biosynthesis in *Klebsiella pneumoniae* clinical isolates. *Microbiology* 155: 4170–4183. doi: [10.1099/mic.0.029017-0](https://doi.org/10.1099/mic.0.029017-0) PMID: [19744990](https://pubmed.ncbi.nlm.nih.gov/19744990/)
36. Lawlor MS, Hsu J, Rick PD, Miller VL (2005) Identification of *Klebsiella pneumoniae* virulence determinants using an intranasal infection model. *Mol Microbiol* 58: 1054–1073. doi: [10.1111/j.1365-2958.2005.04918.x](https://doi.org/10.1111/j.1365-2958.2005.04918.x) PMID: [16262790](https://pubmed.ncbi.nlm.nih.gov/16262790/)
37. Schneiders T, Amyes SG, Levy SB (2003) Role of AcrR and *ramA* in fluoroquinolone resistance in clinical *Klebsiella pneumoniae* isolates from Singapore. *Antimicrob Agents Chemother* 47: 2831–2837. doi: [10.1128/AAC.47.9.2831-2837.2003](https://doi.org/10.1128/AAC.47.9.2831-2837.2003) PMID: [12936981](https://pubmed.ncbi.nlm.nih.gov/12936981/)
38. Veleba M, Higgins PG, Gonzalez G, Seifert H, Schneiders T (2012) Characterization of RarA, a novel AraC family multidrug resistance regulator in *Klebsiella pneumoniae*. *Antimicrob Agents Chemother* 56: 4450–4458. doi: [10.1128/AAC.00456-12](https://doi.org/10.1128/AAC.00456-12) PMID: [22644028](https://pubmed.ncbi.nlm.nih.gov/22644028/)
39. Martin RG, Gillette WK, Rhee S, Rosner JL (1999) Structural requirements for marbox function in transcriptional activation of *mar/sox/rob* regulon promoters in *Escherichia coli*: sequence, orientation and spatial relationship to the core promoter. *Mol Microbiol* 34: 431–441. doi: [10.1046/j.1365-2958.1999.01599.x](https://doi.org/10.1046/j.1365-2958.1999.01599.x) PMID: [10564485](https://pubmed.ncbi.nlm.nih.gov/10564485/)
40. Martin RG, Gillette WK, Rosner JL (2000) Promoter discrimination by the related transcriptional activators MarA and SoxS: differential regulation by differential binding. *Mol Microbiol* 35: 623–634. doi: [10.1046/j.1365-2958.2000.01732.x](https://doi.org/10.1046/j.1365-2958.2000.01732.x) PMID: [10672184](https://pubmed.ncbi.nlm.nih.gov/10672184/)
41. Ricci V, Tzakas P, Buckley A, Piddock LJ (2006) Ciprofloxacin-resistant *Salmonella enterica* serovar Typhimurium strains are difficult to select in the absence of AcrB and TolC. *Antimicrob Agents Chemother* 50: 38–42. doi: [10.1128/AAC.50.1.38-42.2006](https://doi.org/10.1128/AAC.50.1.38-42.2006) PMID: [16377664](https://pubmed.ncbi.nlm.nih.gov/16377664/)
42. Hobbs EC, Yin X, Paul BJ, Astarita JL, Storz G (2012) Conserved small protein associates with the multidrug efflux pump AcrB and differentially affects antibiotic resistance. *Proc Natl Acad Sci U S A* 109: 16696–16701. doi: [10.1073/pnas.1210093109](https://doi.org/10.1073/pnas.1210093109) PMID: [23010927](https://pubmed.ncbi.nlm.nih.gov/23010927/)
43. Padilla E, Llobet E, Domenech-Sanchez A, Martinez-Martinez L, Bengoechea JA, et al. (2010) *Klebsiella pneumoniae* AcrAB efflux pump contributes to antimicrobial resistance and virulence. *Antimicrob Agents Chemother* 54: 177–183. doi: [10.1128/AAC.00715-09](https://doi.org/10.1128/AAC.00715-09) PMID: [19858254](https://pubmed.ncbi.nlm.nih.gov/19858254/)
44. Raetz CR, Reynolds CM, Trent MS, Bishop RE (2007) Lipid A modification systems in gram-negative bacteria. *Annu Rev Biochem* 76: 295–329. doi: [10.1146/annurev.biochem.76.010307.145803](https://doi.org/10.1146/annurev.biochem.76.010307.145803) PMID: [17362200](https://pubmed.ncbi.nlm.nih.gov/17362200/)

45. Velkov T, Soon RL, Chong PL, Huang JX, Cooper MA, et al. (2013) Molecular basis for the increased polymyxin susceptibility of *Klebsiella pneumoniae* strains with under-acylated lipid A. *Innate Immun* 19: 265–277. doi: [10.1177/1753425912459092](https://doi.org/10.1177/1753425912459092) PMID: [23008349](https://pubmed.ncbi.nlm.nih.gov/23008349/)
46. Needham BD, Trent MS (2013) Fortifying the barrier: the impact of lipid A remodelling on bacterial pathogenesis. *Nat Rev Microbiol* 11: 467–481. doi: [10.1038/nrmicro3047](https://doi.org/10.1038/nrmicro3047) PMID: [23748343](https://pubmed.ncbi.nlm.nih.gov/23748343/)
47. Gunn JS, Ryan SS, Van Velkinburgh JC, Ernst RK, Miller SI (2000) Genetic and functional analysis of a PmrA-PmrB-regulated locus necessary for lipopolysaccharide modification, antimicrobial peptide resistance, and oral virulence of *Salmonella enterica* serovar typhimurium. *Infect Immun* 68: 6139–6146. doi: [10.1128/IAI.68.11.6139-6146.2000](https://doi.org/10.1128/IAI.68.11.6139-6146.2000) PMID: [11035717](https://pubmed.ncbi.nlm.nih.gov/11035717/)
48. Buckley AM, Webber MA, Cooles S, Randall LP, La Ragione RM, et al. (2006) The AcrAB-TolC efflux system of *Salmonella enterica* serovar Typhimurium plays a role in pathogenesis. *Cell Microbiol* 8: 847–856. doi: [10.1111/j.1462-5822.2005.00671.x](https://doi.org/10.1111/j.1462-5822.2005.00671.x) PMID: [16611233](https://pubmed.ncbi.nlm.nih.gov/16611233/)
49. Mduli KE, Witte PR, Kline T, Barb AW, Erwin AL, et al. (2006) Molecular validation of LpxC as an anti-bacterial drug target in *Pseudomonas aeruginosa*. *Antimicrob Agents Chemother* 50: 2178–2184. doi: [10.1128/AAC.00140-06](https://doi.org/10.1128/AAC.00140-06) PMID: [16723580](https://pubmed.ncbi.nlm.nih.gov/16723580/)
50. Moreira CG, Herrera CM, Needham BD, Parker CT, Libby SJ, et al. (2013) Virulence and stress-related periplasmic protein (VisP) in bacterial/host associations. *Proc Natl Acad Sci U S A* 110: 1470–1475. doi: [10.1073/pnas.1215416110](https://doi.org/10.1073/pnas.1215416110) PMID: [23302685](https://pubmed.ncbi.nlm.nih.gov/23302685/)
51. Moranta D, Regueiro V, March C, Llobet E, Margareto J, et al. (2010) *Klebsiella pneumoniae* capsule polysaccharide impedes the expression of beta-defensins by airway epithelial cells. *Infect Immun* 78: 1135–1146. doi: [10.1128/IAI.00940-09](https://doi.org/10.1128/IAI.00940-09) PMID: [20008534](https://pubmed.ncbi.nlm.nih.gov/20008534/)
52. Croucher NJ, Fookes MC, Perkins TT, Turner DJ, Marguerat SB, et al. (2009) A simple method for directional transcriptome sequencing using Illumina technology. *Nucleic Acids Res* 37: e148. doi: [10.1093/nar/gkp811](https://doi.org/10.1093/nar/gkp811) PMID: [19815668](https://pubmed.ncbi.nlm.nih.gov/19815668/)
53. Quail MA, Kozarewa I, Smith F, Scally A, Stephens PJ, et al. (2008) A large genome center's improvements to the Illumina sequencing system. *Nat Methods* 5: 1005–1010. doi: [10.1038/nmeth.1270](https://doi.org/10.1038/nmeth.1270) PMID: [19034268](https://pubmed.ncbi.nlm.nih.gov/19034268/)
54. Briggs GS, Mahdi AA, Wen Q, Lloyd RG (2005) DNA binding by the substrate specificity (wedge) domain of RecG helicase suggests a role in processivity. *J Biol Chem* 280: 13921–13927. doi: [10.1074/jbc.M412054200](https://doi.org/10.1074/jbc.M412054200) PMID: [15695524](https://pubmed.ncbi.nlm.nih.gov/15695524/)
55. Schneiders T, Levy SB (2006) MarA-mediated transcriptional repression of the *rob* promoter. *J Biol Chem* 281: 10049–10055. doi: [10.1074/jbc.M512097200](https://doi.org/10.1074/jbc.M512097200) PMID: [16478729](https://pubmed.ncbi.nlm.nih.gov/16478729/)
56. Barbosa TM, Levy SB (2002) Activation of the *Escherichia coli nfnB* gene by MarA through a highly divergent marbox in a class II promoter. *Mol Microbiol* 45: 191–202. doi: [10.1046/j.1365-2958.2002.03006.x](https://doi.org/10.1046/j.1365-2958.2002.03006.x) PMID: [12100559](https://pubmed.ncbi.nlm.nih.gov/12100559/)
57. Cano V, Moranta D, Llobet-Brossa E, Bengoechea JA, Garmendia J (2009) *Klebsiella pneumoniae* triggers a cytotoxic effect on airway epithelial cells. *BMC Microbiol* 9: 156. doi: [10.1186/1471-2180-9-156](https://doi.org/10.1186/1471-2180-9-156) PMID: [19650888](https://pubmed.ncbi.nlm.nih.gov/19650888/)
58. Regueiro V, Campos MA, Pons J, Alberti S, Bengoechea JA (2006) The uptake of a *Klebsiella pneumoniae* capsule polysaccharide mutant triggers an inflammatory response by human airway epithelial cells. *Microbiology* 152: 555–566. doi: [10.1099/mic.0.28285-0](https://doi.org/10.1099/mic.0.28285-0) PMID: [16436443](https://pubmed.ncbi.nlm.nih.gov/16436443/)
59. George AM, Hall RM, Stokes HW (1995) Multidrug resistance in *Klebsiella pneumoniae*: a novel gene, *ramA*, confers a multidrug resistance phenotype in *Escherichia coli*. *Microbiology* 141: 1909–1920. doi: [10.1099/13500872-141-8-1909](https://doi.org/10.1099/13500872-141-8-1909) PMID: [7551053](https://pubmed.ncbi.nlm.nih.gov/7551053/)